

## 2-color STED CW Sample Preparation

The protocol doesn't differ from any other normal immunocytochemistry, and we are encouraging you to stay as close as possible to your usual immunocytochemistry protocol. Only exchange the secondary antibody (if necessary) and test different concentrations for optimization of the signal.

For cell cultures, please grow your cells on a **#1.5 coverslip (0.170 mm)**, or on a **cell culture chamber slide with a #1.5 glass bottom**. **Do NOT use any other coverslips thickness** as microscope objectives are corrected for # 1.5 coverslips and therefore best penetration depth will be obtained with these coverglasses. We recommend the Lab-Tek™ II Chambered Coverglass products with **#1.5 Borosilicate** Coverglass (no commercial interest).

If you are using tissue sections, please use a # 1.5 coverslip to mount your sections.

Prepare your sample as usual with your primary antibody and the typical washing steps as well as the preincubation in PBS/normal serum/ permeabilisant (Triton X100 or other).

The **choice and combination** of the fluorescent secondary antibodies (or fluorescence protein) is important in order to be able to image your sample in STED mode using the Leica TCS STED CW.

For **single color STED CW**, the following dyes has been tested and reported to give excellent results in STED CW:

- DyLight 488
- Chromeo 505
- Oregon green 488
- Alexa 488 , Alexa 514.

For **2-color STED CW**, the following dyes have been tested and showed good results in combination with one dye from the single color STED CW list.:

- Atto 425, antibody or Streptavidin conjugated
- Pacific Orange, antibody conjugated

## Dye conjugates:

In the following tables show the *recommended dyes and dye combinations which gave good results.*

### Recommended pair for 2-color STED CW

<u>Channel 1</u>		<u>Channel 2</u>	<u>Comment</u>
Pacific Orange (Secondary Antibodies conjugated)	+	DyLight488 / Alexa488 (excluding Oregon Green 488 and Chromeo 505)	Dye separation required
Atto 425 (Anti-mouse)	+	DyLight 488, Chromeo 505, Oregon Green 488, Alexa 488	To be used with protein or structure of interest bigger than 100 nm due to a lower efficiency of the dye in STED CW. <b>No dye separation required.</b>
NBD-X	+	Any other green dye reported to work for STED CW	No commercial conjugates for NBD-X so far

### Recommended concentration and possible providers (no commercial interest):

*We encourage people to adjust the concentration as needed*

#### **Channel 1:**

- Atto 425 Goat  $\alpha$ -mouse (2-4  $\mu$ g/ml; *On Demand, contact Myriam*)
- Pacific Orange (1:200; *Invitrogen*),  $\alpha$ -mouse (P-31585) or  $\alpha$ -rabbit (P-31584).

#### **Channel 2:**

- DyLight 488 (1-2  $\mu$ g/ml, *Rockland Immuno*).Goat  $\alpha$ -mouse (610-141-121) and  $\alpha$ -rabbit (611-141-122)
- Oregon Green 488 (1:200; *Invitrogen*), Goat  $\alpha$ -mouse (O-11033) or  $\alpha$ -rabbit (O-11038).
- Alexa 488 (1:200 to 1:400).
- Atto 488 (1:1000).
- Chromeo 488 (1:100 to 1:200).

**Fluorescent Proteins:**

<b><u>Channel 1 (458 ex)</u></b>	<b><u>Channel 2 (514 ex)</u></b>
mCerulean	mCitrine, mVenus
	<b>EmeraldGFP(EmGFP), eGFP</b>
	eYFP, mYFP

This list is not exhaustive and other secondary antibodies or FPs can be tried in STED mode as long as ***they can be excited with the 458, 488 or 514 nm lasers AND have their peak emission wavelength around 510*** (as well as to have a tail emission at 592 nm).

**Do Not use List:**

<b>Do NOT Use</b>
<ul style="list-style-type: none"> <li>Other Fluorescence Proteins <b><u>not</u> excitable by 458, 488 or 514 nm lasers.</b></li> </ul>
<ul style="list-style-type: none"> <li>Do <b>NOT</b> use <b>DAPI</b></li> </ul>
<ul style="list-style-type: none"> <li>Do <b>NOT</b> use <b>QDOTs</b></li> </ul>
<ul style="list-style-type: none"> <li>Do <b>NOT</b> use other fluorescence excited with the 405 nm laser</li> </ul>

**Mounting media:**

***If working with cell culture***

Best	Good	Do Not Use
<b>Prolong + Antifade</b> (will polymerize)	<b>Mowiol + Antifade</b> (will polymerize)	Slowfade
<b>Thiodiethanol</b> (TDE, Sigma, #88559. See below about the concentration ). <b>Add Antifade</b> (will stay viscous)		Vectashield

**Prolong Antifade Kit (Invitrogen P7481, w/o DAPI)**, a mounting which will polymerize, has been tested in STED mode and gave excellent results as long as the protein of interest is located within 30 µm from the coverslip. It will also depend on the tissue. While brain can be imaged quite deep using visible lasers, other denser tissues may be more difficult to image at depth >30 µm. If your target is further away, then TDE is the perfect choice (see below for the concentration and preparation steps).

**Thiodiethanol** (TDE, Sigma, #88559) has been used with excellent results (see reference below) especially for deep imaging. The TDE concentration **must be gradually enhanced to obtain a final refractive index of 1.514 (see addendum page)**, which is reached using a TDE concentration of 97%. If using TDE, the coverslip must be sealed using invisible nail polish or other sealants. Please, be sure that the sealant is not quenching your fluorescence or creating any autofluorescence.

***For tissue sections***

**TDE** is the best selection. As described before, the TDE concentration **must be** gradually enhanced to obtain a final refractive index of 1.514, which is reached using a TDE concentration of 97% (see the reference below). Sequential steps in TDE 50%, 70% (15 minutes at each step), then in 97% + antifade as final mounting media must be undertaken. Here again, if using TDE, the coverslip must be sealed using invisible nail polish or other sealants. Please be sure that the sealant is not quenching your fluorescence or creating any autofluorescence.

**Reference about TDE:**

*Microsc Res Tech. 2007 Jan;70(1):1-9.*

**2,2'-thiodiethanol: a new water soluble mounting medium for high resolution optical microscopy.**

*Staudt T, Lang MC, Medda R, Engelhardt J, Hell SW.*

**Prolong Antifade Kit** can be used too if the structure of interest is close from the coverslip. If you need to image further away from the coverslip, then the TDE should be tried.

### Antifade reagents:

All these mounting media **must have antifade freshly mixed in it. *Do NOT use a solution mixed with any antifade which is older than few hours.***

- **P-phenylenediamine** (PPD, 0.01-0.1%) is probably the most effective antifade, particularly in combination with the TDE.
- The proprietary antifade from Invitrogen supplied in the **Prolong Antifade Kit** special packaging can also be used (**component A**). The antifade is coming separately in small opaque tubes, and therefore may be used with TDE. Just add 1 ml of TDE 97% to the tube (component A), mix well and centrifuge the tube to eliminate as much as possible the bubbles. Pipette at the bottom of the tube avoid any air bubbles left.
- **Most Live Cell experiments usually do not include any adjunction of antifade.** Nevertheless, **N-propyl gallate** (NPG, 2%) while not very soluble, is non-toxic and can be used on live cells. Normally, the recommended medium for the specific cells (without Phenol Red, and buffered at pH 7.4) should be used, or maybe replaced with PBS.
- **DABCO** (2.5%) is a well-known antifade, while not as effective as the PPD, but is also less toxic.

### Counterstain:

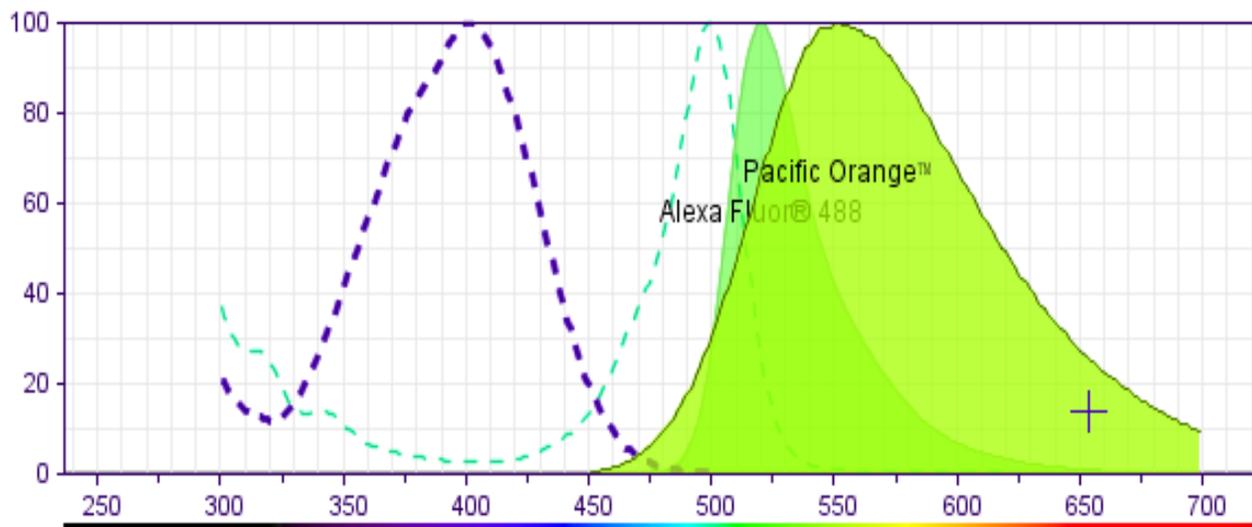
You have the possibility to counterstain and image in confocal mode your STED CW samples using a near far red dye like **Alexa, DyLight or Atto 633, 647 or 655 nm**. If using an orange-red fluorescence like Alexa, DyLight or Atto 543, 555, or 565 nm there is a possibility of crosstalk when using the Argon laser in the range of 488 and 514, and/or strong absorption using the 592 nm STED CW laser. Therefore it would be appropriate to exchange these counterstains to the ones cited above.

**APPENDIX 1:**

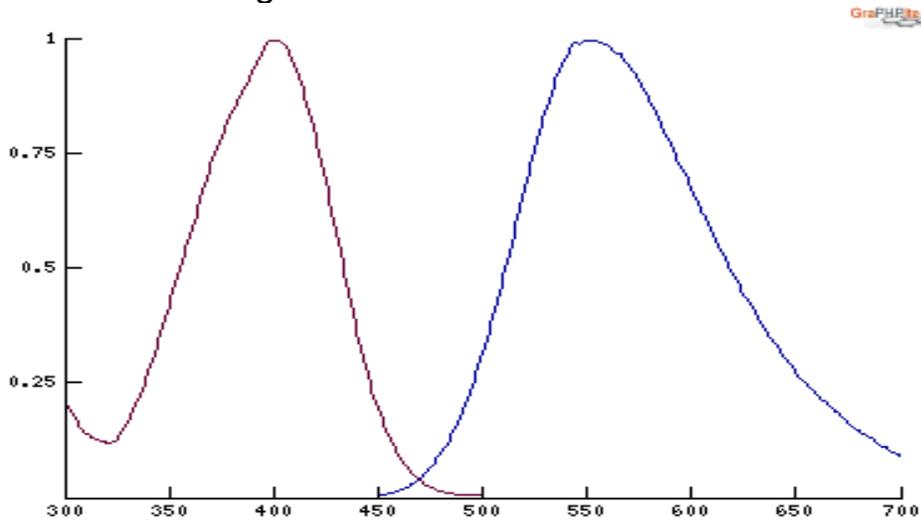
**Examples of optimization of the acquisition for selected dyes pairs, and selected spectra:**

<b>Dye1</b>			<b>+</b>	<b>Dye2</b>		
<b>Name</b>	<b>Excitation</b>	<b>Emission</b>		<b>Name</b>	<b>Excitation</b>	<b>Emission</b>
Atto 425	458	465 - 510	+	Oregon Green / Any 488 / Any 514 dyes	514	520 - 580
Pacific Orange	458	535 - 580	+	DyLight 488, Alexa 488 / Any 488 (excluding Oregon Green 488)	488 496	500 – 530 505 - 530
Pacific Orange	458	500 -580	+	Any 488 (excluding Oregon Green 488)	488 496 514	500 – 580 505 – 580 525 - 580
NBD-X	458	535 – 580 Or 500 - 580	+	DyLight 488, Alexa 488 / Any 488 (excluding Oregon Green 488)	488	500 – 530 Or 500 -580

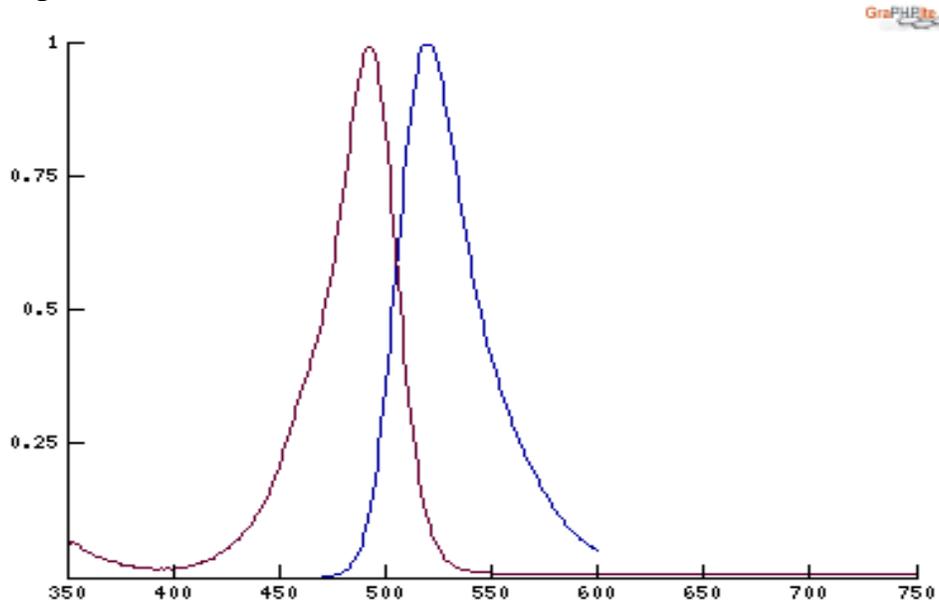
**Pacific Orange + Alexa (or DyLight) 488**



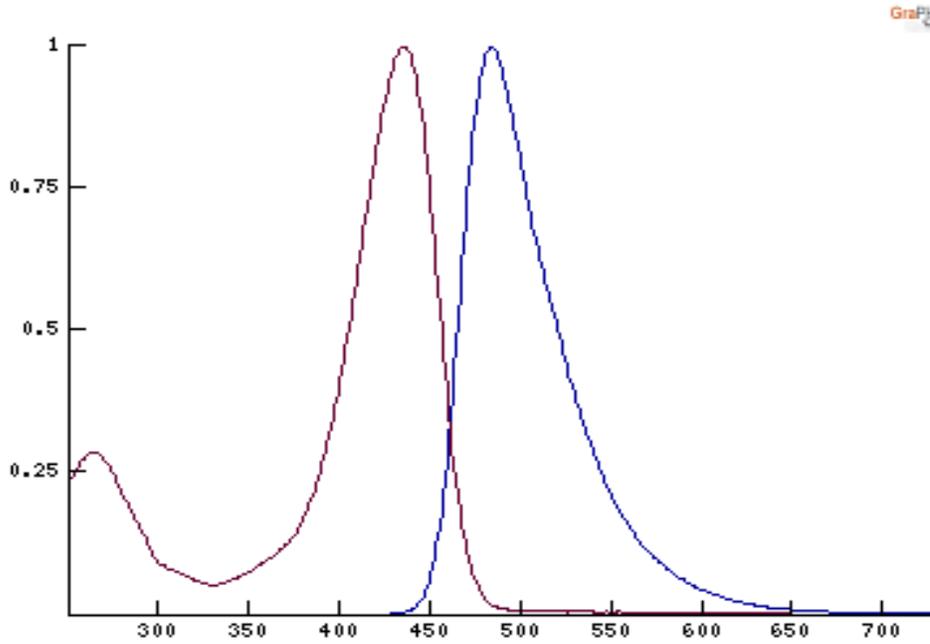
### Pacific Orange



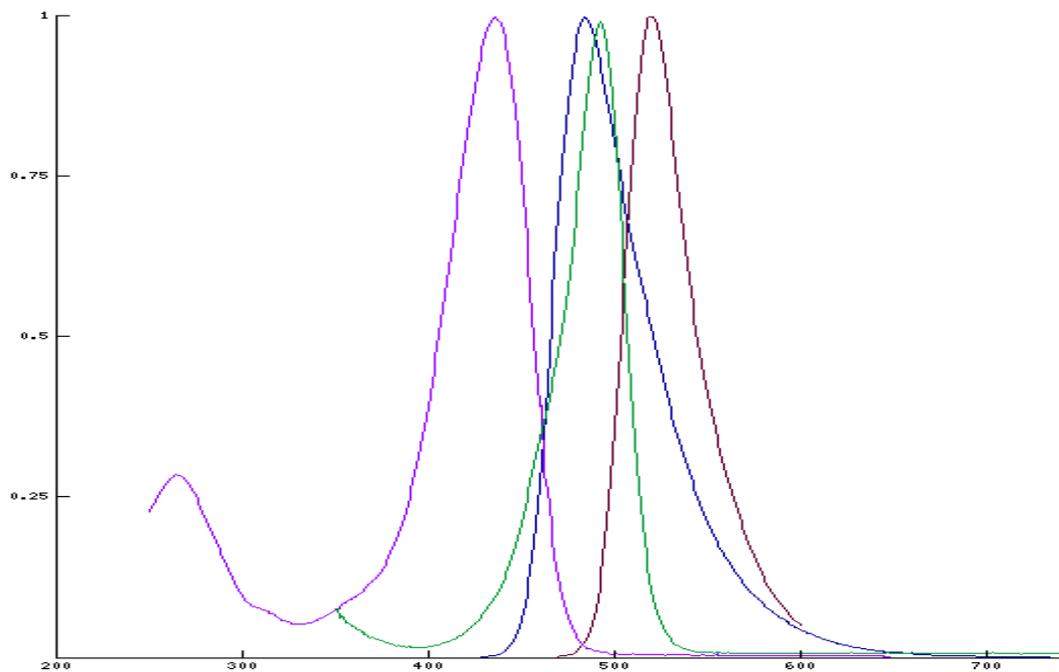
### DyLight 488



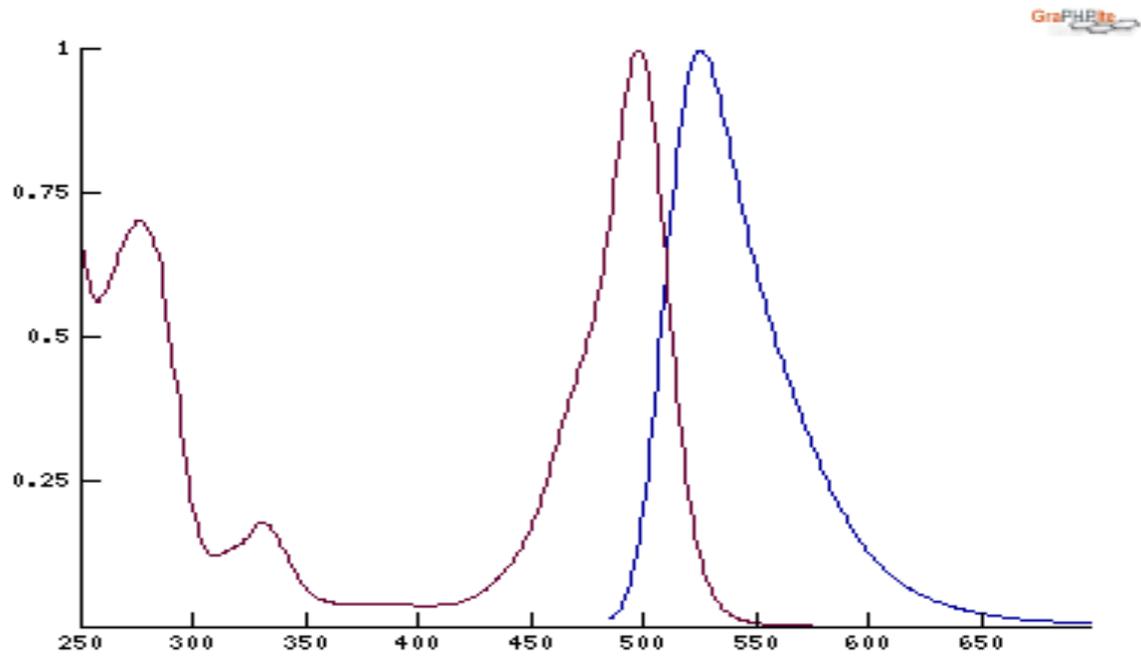
ATTO 425



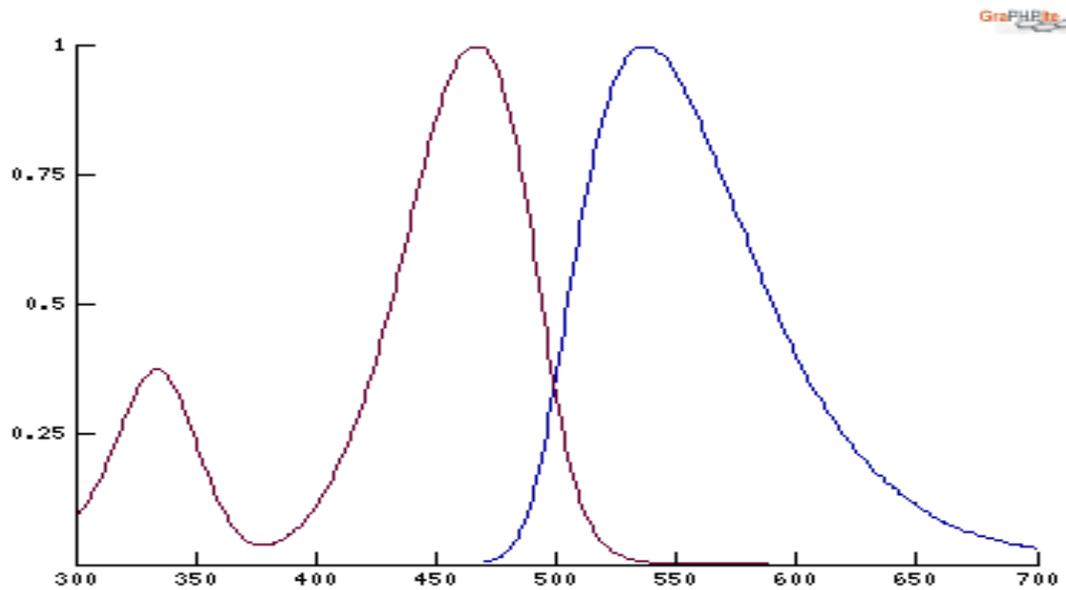
ATTO 425 (Ex / Em) + DyLight 488 (Ex / Em)



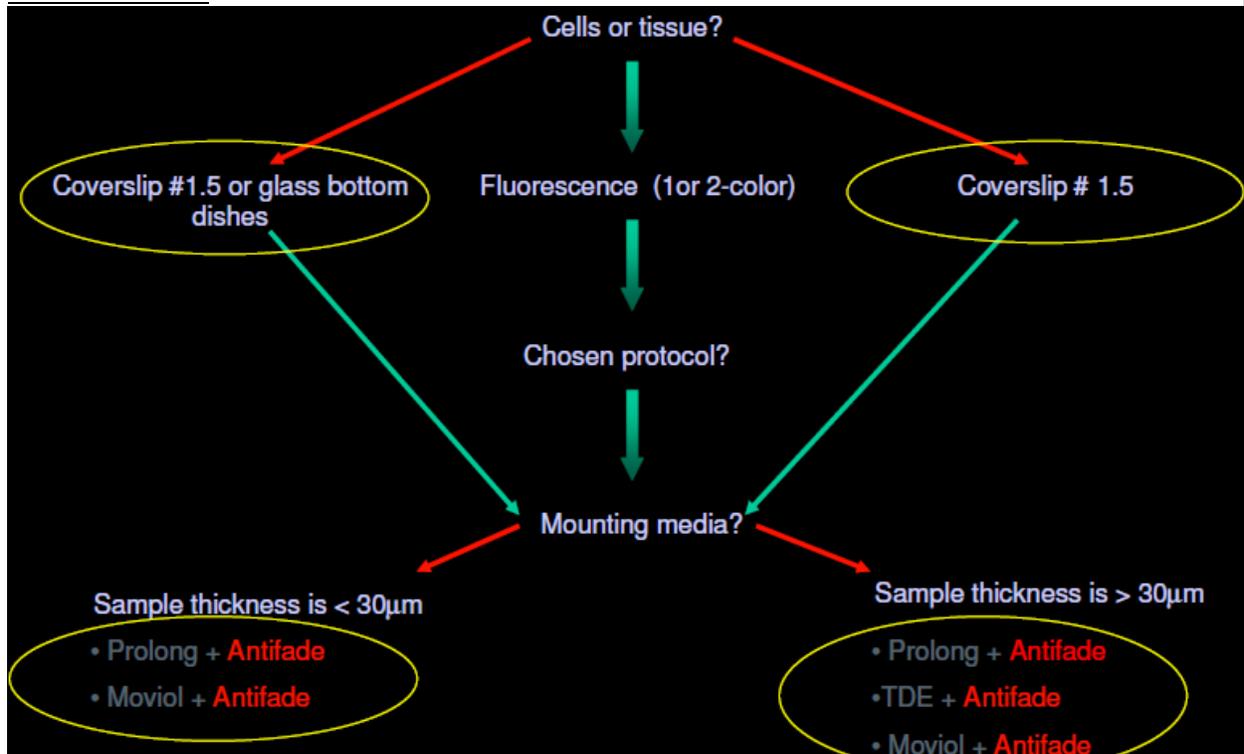
### Oregon Green 488



### NBD-X

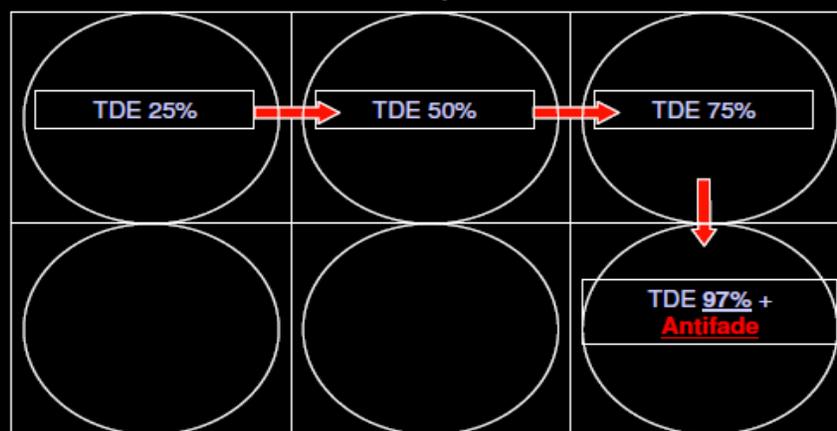


**APPENDIX 2:**



**STED CW: Thiodiethanol preparation for tissue mounting**  
(TDE, Sigma, #88559)

6 Well-plate



- 15-30 minutes incubation at each step (depending on the sections thickness).
- Mix **VERY WELL** the antifade (*pipette in and out*) with the TDE 97% prior to mount the tissue sections (eliminate the bubbles by centrifugation).
- Use nail polish (high quality uncolored nail polish) on the #1.5 coverslip corner to keep it in place for the night (must be kept flat).
- Seal the coverslip edges the following morning with nail polish.

Living up to Life



If you need any help or if you have any questions, please be sure to call me at **866-830-0735, opt 3.**

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