



Purdue University

Veterinary Technology Distance Learning Program

Lynn Hall, Purdue University, West Lafayette, Indiana 47907

Necropsy Clinical Mentorship VM 22400

Criteria Handbook and Log Book

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STUDENT INFORMATION

GOALS OF VM 22400 NECROPSY CLINICAL MENTORSHIP

Working with a veterinary care facility, the student will practice several tasks under the supervision of a clinical mentor (veterinarian or accredited veterinary technician).

In order to achieve the goals for this Clinical Mentorship, the tasks must be performed to the level of competency as outlined by the *Criteria* for each task.

The student is responsible for providing documentation for each task as defined by the *Materials Submitted for Evaluation and Verification* section on each task.

In addition to the documentation, the student will be evaluated by the Clinical Mentorship site supervisor on how well the student met the outlined Criteria for each task.

Final approval of successful performance and completion of the Clinical Mentorship will be made by the Purdue University instructor in charge of the Clinical Mentorship. This approval will be based upon the documentation provided by the student.

The Purdue University instructor in charge has the option to require additional documentation if, in their judgment, the student has not performed and/or documented the task to the level set by the Criteria.

Documentation of completed tasks is essential to validating the educational process and insuring that the performance of graduates of the Veterinary Technology Distance Learning Program meets the standards of quality required by the Purdue University School of Veterinary Medicine faculty and the American Veterinary Medical Association accrediting bodies.

CONTACT PERSON

Any questions regarding the Clinical Mentorship process should be directed to:

Pam Phegley, BS, RVT
1240 Lynn Hall
Purdue University
West Lafayette, IN 47907
(765) 496-6809
phegley@purdue.edu

PRE-REQUISITES FOR VM 22400 NECROPSY CLINICAL MENTORSHIP

Course Pre-requisites

VCS 23100	Small Animal Nursing and Health Management IV
VCS 14900	Large Animal Nursing and Health Management II
VPB 22600	Parasitology II
VPB 22700	Microbiology
VPB 24100	Public and Occupational Health II

Contracts and Agreements

Because of legal, liability and AVMA accreditation issues, the following documents must be completed prior to beginning the Clinical Mentorship

1. Clinical Mentorship Site Facility Requirements Agreement
2. Clinical Mentorship Agreement
3. Clinical Mentorship Supervisor Agreement
4. Health Risk and Insurance Acknowledgment
5. Professional Liability Insurance Coverage
6. Agreement and Release of Liability
7. Technical Standards Acknowledgment

These forms are available on the VT-DLP web site for downloading, printout, and completion, or by phone request from the VT-DLP office (765-496-6579).

If more than one Clinical Mentorship course is taken, a separate Clinical Mentorship Site Facility Requirements Agreement, Clinical Mentorship Agreement, and Clinical Mentorship Supervisor Agreement must be completed for each course.

Failure to complete and return the listed documents and the payment for Student Professional Liability Insurance Coverage will prevent the student from enrolling in the Clinical Mentorship.

Insurance

Two types of insurance are recommended or required for the student working in a Clinical Mentorship.

Health Insurance is highly recommended to cover the medical expenses should the student become injured while on the job. It is the student's responsibility to procure such insurance.

Liability insurance is required to protect the student in the event of a suit filed against the student for acts he/she performed while in the Clinical Mentorship. Each VT-DLP student is required to purchase, for a nominal fee, Professional Liability Insurance through Purdue University. This is done by completing the Professional Liability Insurance Coverage form and sending a check for the fee. The fee covers from the time of initiation of coverage until the subsequent May 31st.

Students will not be enrolled in Clinical Mentorships until the Professional Liability Insurance is paid, and the student is covered by the policy.

SELECTING THE CLINICAL MENTORSHIP SITE – FACILITY REQUIREMENTS

You must visit the Clinical Mentorship Site and determine if the following equipment is readily available to you for use during your Clinical Mentorship. You must complete and have the facility veterinarian sign the Clinical Mentorship Site Facility Requirements Agreement.

The veterinary care facility must be equipped:

With the following equipment:

- Necropsy knife (sturdy and able to be sharpened)
- Scalpel handle
- Mayo or Metzenbaum scissors
- Forceps
- Serrated utility scissors
- Pruning shears for other instrument for cutting ribs

With the following items:

- Heavy-duty latex gloves
- Scalpel blades
- Jars of formalin
- Sterile culture tubes or culture swabs
- Zipper-top plastic bags
- Indelible marking pen

SELECTION OF THE CLINICAL MENTORSHIP SUPERVISOR

The Clinical Mentorship Supervisor is the person who will sign your Logbook and assess your performance at the Clinical Mentorship site. This person must be a credentialed veterinary technician (graduated from an AVMA accredited program and/or met State requirements for credentialing as a veterinary technician) or a licensed veterinarian.

An individual who claims to be a “veterinary technician” but has not met the criteria for credentialing above is not eligible to be a mentorship supervisor.

The individual is not considered to be an employee of Purdue University when acting as your Clinical Mentorship supervisor.

The Clinical Mentorship Supervisor must complete the *Clinical Mentorship Supervisor Agreement*. You must return this agreement with the other agreements prior to beginning your Clinical Mentorship.

Should your Clinical Mentorship Supervisor change during the course of the Clinical Mentorship, you will need to have your new supervisor complete a *Clinical Mentorship Supervisor Agreement* and return it to the Purdue VT-DLP office. These forms are available on the VT-DLP web site for downloading and printing.

CRITERIA HANDBOOK AND LOG BOOK

This Criteria Handbook and Log Book contains the list of tasks that must be successfully completed in order to receive credit for this Clinical Mentorship. You are expected to have learned the basics of how, why, and when each procedure is to be done from the courses listed as pre-requisites for this Clinical Mentorship. This booklet contains the directions and forms that must be followed and completed in order to meet the standards set for successful completion of this Clinical mentorship.

Please read each component of each task carefully before doing the task to minimize the number of times you have to repeat the task. The components of each task are summarized:

Goal – Describes the ultimate outcome of the task you will perform.

Description – Lists the physical acts that you will perform, and under what conditions these acts will be completed.

Criteria – Lists *specific, observable, objective* behaviors that you must demonstrate for each task. Your ability to demonstrate *each* of these behaviors will be required in order to be considered as having successfully completed each task.

Number of Times Task Needs to be Successfully Performed – States the required number of times to repeat the tasks. The patient's name and the date each repetition of task was performed must be recorded by the Clinical Mentorship Supervisor.

Materials Submitted for Evaluation and Verification – These specific materials, which usually include some video or photographic materials, must be submitted to demonstrate that you actually performed the task as stated. Each evaluation states specifically what must be shown in the submitted materials.

The Purdue University course instructor for this Clinical Mentorship has the option to request further documentation if the submitted materials do not clearly illustrate the required task.

It is recommended that the video materials document all angles of the procedure. The purpose of the video and photographic material is to provide "concrete evidence" that you were able to perform the task to the standard required.

If you do not own a video camera, they can usually be borrowed or rented. Pre-planning the video procedures will help reduce the need to redo the video documentation. Feel free to explain what you are doing as you perform for the video documentation. Sometimes, voiceovers may need to be done to clearly explain what task is being performed.

Videotapes, pictures, the Criteria Handbook and Logbook and any other required documentation will not be returned. These items will be kept at Purdue as documentation of the student's performance for accreditation purposes.

This validation is essential to help the Purdue VT-DLP meet AVMA accreditation criteria. Therefore, it is essential that you follow the evaluation and validation requirements.

Summary Evaluation Forms – Each task has a form that must be completed and signed by the Clinical Mentorship Supervisor.

Supplementary Materials – Logs, written materials, photographs, or other forms/ documentation may be required for specific tasks. Be sure to read the Materials Submitted for Evaluation section very carefully and return all documented evidence as prescribed.

COMPLETION OF THE CLINICAL MENTORSHIP

The Clinical Mentorships are designed to follow the semester format of Purdue University. However, you may complete the Clinical Mentorship and submit materials any time prior to the end of the semester.

If you are unable to complete the Clinical Mentorship by the end of the semester deadline, you must contact the course instructor for the Clinical Mentorship, the VT-DLP office (765) 496-6579, or Pam Phegley, BS, RVT (phegley@purdue.edu, 765-496-6809) to request an "Incomplete grade" for the semester. The Clinical Mentorship will be treated by the University as it would any other Incomplete graded course. See the VT-DLP Student Handbook for specific information on Incompletes.

When you have completed all of the tasks and the documentation, send the complete compilation of materials to:

Vet Tech Distance Learning
Clinical Mentorship Coordinator
1240 Lynn Hall, Purdue University
West Lafayette, IN 47907

You will be contacted after the materials have been reviewed. The Purdue University instructor in charge has the option to require additional documentation if, in their judgment, the student has not performed or documented the task to the level set by the Criteria.

If additional documentation is deemed necessary by the course instructor or the Clinical Mentorship Coordinator, the student will be contacted and the additional required documentation explained.

Final approval of successful performance and completion of the Clinical Mentorship will be made by the Purdue University instructor in charge of the Clinical Mentorship based upon the documentation provided by the student.

Upon successful completion of the documentation, a grade for the course will be assigned by the course instructor based upon the Clinical Supervisor's evaluations and the documented performance of the tasks.

Notes to Student

As there is no didactic course to accompany the Necropsy Clinical Mentorship, hand-outs have been included in the mentorship logbook to inform the student of techniques and procedures that will be needed to complete the tasks. The first is the technique used by the Purdue Animal Disease Diagnostic Laboratory (ADDL) for performance of necropsy. The second is a page of guidelines for packing and shipping samples to outside labs.

The student should review anatomy to be able to identify anatomical landmarks mentioned in the necropsy technique. In addition, the student should understand all the terminology used to describe the technique or lesions observed (i.e., “transudate”, “in situ”, etc.)

The student should also read pages 127-141 in the 5th edition McCurnin book.

**Thoroughly review and understand the technique
prior to beginning the first necropsy.**

VETERINARY NECROPSY TECHNIQUE

Purdue University
Animal Disease Diagnostic Laboratory

The following is a brief outline of the technique that is to be used when performing postmortem examinations. Do not begin the necropsy until a permission sheet signed by the owner is in your possession. A signed owner's release form must accompany any animal to be euthanized.

Remember that the entire carcass, including all systems and organs, must be carefully examined. Lesions may appear anywhere and care should be taken to expose and examine all lesions. Examine each of the paired organs.

Every animal should be weighed and/or measured (i.e., crown-rump length for aborted feti) prior to prosection.

Preliminary Review and Observations

1. Signalment – species, breed, sex, sexual status, age, color
2. History and clinical diagnosis
3. Clinical pathology
4. External appearance
 - Body condition (adequate or inadequate fat stores, emaciated, etc)
 - Mucous membranes
 - Body orifices
 - General conformation
 - Superficial lesions (tumors, dermatitis, etc.)
 - Hair coat
 - Parasites
 - Lips, gums, cheeks, teeth

Opening the Body Cavities

1. Place animal carcass in lateral recumbency and incise skin at axilla
 - Continue the ventral midline skin incision anteriorly to the symphysis of the mandible and posteriorly to the perineum
 - Do not damage the udder
 - To avoid cutting hair, incise the skin from the subcutaneous side
2. Raise the front leg and scapula and dissect and reflect dorsally
 - Remove the remaining skin between the excised front and rear limb to the level of the spinal column and reflect dorsally
 - Examine the exposed superficial lymph nodes and jugular veins
3. Excise through the “up” rear limb (at the level of the pelvis) and continue to incise through the coxofemoral joint and reflect the rear limb dorsally

4. Examination of the mammary glands or testes
 - Mammary glands and mammary lymph nodes are completely cut away from the body
 - Examine for symmetry, swellings, tumors, atrophy
 - Examine the lymph nodes and incise them
 - Incise the gland through the cistern and teat canal, examining each portion
 - Palpate for thickenings, fibrosis, tumors
5. Examine prepuce and penis
6. Make a paracostal incision through the abdominal wall just behind and parallel to the last rib
 - Extend the incision dorsally to the vertebrae and ventrally to the midline
 - Raise the body wall to avoid cutting viscera
7. Make a paralumbar incision through the abdominal wall caudally to the pelvis
 - Reflect the muscle wall ventrally and expose the abdominal cavity
8. Cut the diaphragm on the right side in an arc from the sternum along its costal attachments to the vertebral column
 - Listen for an in rush of air indicative of negative pressure in the pleural cavity
9. Sever the ribs at their sternal and vertebral ends with a pruning shear or other suitable instrument and lift off the thoracic wall, thus exposing the entire thoracic cavity

Gross Examination of the Thoracic and Abdominal Cavities

1. Examine both cavities and all contents carefully with minimal movement of the viscera
 - Note transudates, exudates, and hemorrhage
 - Open the pericardial sac
 - Note amount, color, and consistency of abnormal fluid accumulations
 - Examine for adhesions, displacements, absence of organs, and size and symmetry of organs in situ
 - Record lesions of organs and perform detailed examination of organs prior to removal
 - Take initial samples for microbiology, especially exudates in body cavities

Examination of the Thoracic Viscera

1. Separate the mandibles at the symphysis
 - Cut along the lingual surface of both sides of the mandible
 - Remove the tongue and pull it down between the rami
 - Disarticulate the hyoid bones. The tongue, larynx, trachea and esophagus are dissected ventrally back to the thoracic inlet

- Lift up viscera and detach heart and lungs from the body wall by cutting dorsal and ventral mediastinum
 - Sever the aorta, post cava and esophagus back to about 2-3 cm anterior to the diaphragm
 - Sever and remove the thoracic viscera (“pluck”)
2. Examine thyroid, parathyroid, and thymus glands
 - Note size, shape, consistency
 - Incise glands examining for lesions
 3. Arrange the organs in approximately normal position
 - Examine tongue by incising transversely
 - Open esophagus and examine carefully
 - Examine bronchial lymph nodes by palpating and incising
 - Observe and palpate lungs for consolidation, emphysema or other abnormal consistency
 - Open the larynx, trachea, bronchi and small bronchioles
 - Note exudates, hemorrhage, foreign bodies or lung worms in bronchial tree
 - Examine areas of consolidation and other abnormal lung tissue by incising
 4. Examine the heart
 - Observe any disproportion of parts (dilation, hypertrophy, anomalies) and alterations in shape; note presence of normal adipose tissue
 - Open heart
 - Cut through the right atrial free wall (including the auricle) horizontally
 - Examine the endocardium and vena cava
 - Examine the atrial side of the right A-V valve
 - Check for sufficiency of valve if indicated
 - Cut through the right A-V valve and wall of the right ventricle, keeping the incision near the interventricular septum
 - Continue the incision around the right ventricle through the pulmonic valve and pulmonary artery
 - Examine for patent ductus arteriosus
 - Open the left atrium and examine in the same manner as the right atrium
 - Cut through the left A-V valve, incising the ventricle through the mid-portion of the free wall
 - Continue the incision to the apex
 - Make a horizontal incision in the ventricle approximately mid-way between the coronary groove and the apex, incising from the first cut to the septum
 - At the septum, cut upward through the aortic valve and aorta
 - This process should result in a small flap of left heart with aortic valve on one side and left A-V valve on the other
 - Examine vessels, valves and septa for anomalies
 - Examine endocardium and myocardium

Examination of Abdominal Viscera

1. Remove the spleen; examine grossly and incise several times
2. Examine the pancreas grossly
3. Make a small incision into the duodenum at the level of the pancreatic duct and apply manual pressure to the gall bladder to see if bile enters the intestine
4. Remove and examine the liver
 - Examine the peritoneal surface for fibrosis or adhesions
 - Excise the liver from the diaphragm
 - Note the size, shape, weight, color and consistency
 - Open the gall bladder and the larger bile ducts
 - Examine for stones, inflammation, flukes, thickening of the wall
 - Palpate and incise the liver liberally from the abdominal surface; observe for necrosis, fibrosis, abscesses, etc.
5. Examine the adrenal glands (prior to removing the kidneys)
 - Cut adrenals in cross-section and note cortical-medullary ratio
6. Remove urinary organs as a unit, including both kidneys, ureters and urinary bladder
 - Cut each kidney longitudinally in half from the convex surface to the hilus and note alterations in color, consistency, size, etc.
 - Strip off capsule and examine the kidney surface
 - Note the ease with which the capsule comes off
 - Open and inspect the ureters, bladder and urethra
 - Inspect all mucous and serous surfaces
 - Open vagina, cervix and uterine horns along their dorsal borders and examine carefully all surfaces
 - Examine ovaries for cysts, corpora lutea, atrophy, etc.
 - Examine male accessory sex organs; observe size, consistency, inflammation, etc.
7. Remove the stomach and intestines to the rectum
 - Place the rectum over the lumbar area when it is cut so that the abdomen will not be contaminated
 - Free the intestine from the mesentery as it is removed and observe its lymph nodes

The examination of the gastrointestinal tract is delayed until last so that instruments and other tissues are not contaminated by gastrointestinal flora

Examination of the Musculoskeletal System

1. Open the stifle, hock and humero-scapular joints
 - To open the stifle, cut the straight patellar ligament 1/3 of the way proximal to the tibial tuberosity and medial to the trochlea of the femur, and reflect the patella
 - Observe synovia, articular surfaces, articular cartilages, and synovial membranes

2. Examination of the muscular system
 - Examine and incise the muscles of various parts of the body, especially lumbar and thigh muscles; check development, color, etc.
3. Examination of the skeletal system
 - Examine body for broken bones or healed fractures
 - For marrow inspection, remove femoral head with shears and crack femur longitudinally

Examination of the Eyes

1. Remove the eyeball from the orbit if indicated (not routine)
 - Incise periorbital tissues and avoid direct contact with the eye
 - Look for corneal opacities, cataracts, tumors, etc.

Examination of the Central Nervous System

1. Remove the head from the body at the atlanto-occipital articulation
 - Incise the spinal cord before excessive traction is placed on the skull
2. Reflect skin and muscles of the head and examine skull for traumatic lesions
3. Remove the brain as described below
 - Make a transverse cut behind the orbits (exact location varies in species) using a hacksaw
 - Make lateral cuts from the ends of the transverse cuts just medial to the occipital condyles (leave room for brain to be removed intact)
 - Lift off bony cap carefully with a chisel
 - Incise the dura over the dorsal brain surface and incise the tentorium cerebelli
 - Hold the skull with the nose pointing upward and tap it gently on the table
 - o Carefully cut the olfactory tracts and other cranial nerves and allow the brain to slip out
 - o Avoid traction on the brain
 - Remove the pituitary gland by cutting diaphragmatic sella on both sides, clipping the bony projection posterior to the gland, and cutting soft tissues around the gland with scissors
4. Observe the dura
5. Incise the brain transversely (1-cm slices) and look for lesions
 - When entire brain is to be fixed, make only one transverse cut into lateral ventricles so fixative may enter tissues

Examination of the Gastrointestinal Tract

1. The esophagus has been opened
2. Open the stomach along the greater curvature
 - Observe the mucosal and serosal surfaces; ingesta must be removed
 - Examine for hemorrhage, parasites, foreign bodies, abnormal ingesta, etc.
3. Open the small intestine
 - Observe all surfaces and ingesta
 - Leave 1-inch segments closed for histopathology
4. Open the cecum and colon back to the anus, and examine carefully

Species-Specific Procedures

1. Horse
 - When the abdomen is opened, move the left parts of the large colon cranially so that the pelvic flexure is lying anterior; move the cecum dorsocranially, the small intestine over the right flank, and the small colon posterior and down
 - The mucosa of the guttural pouches is examined when the head is disarticulated
 - The cranial mesenteric artery should be opened from the aorta past the ileal-cecal and colic artery bifurcations
2. Ruminants
 - When the abdomen is opened, place small intestine and colon over the right lumbar area; examine the forestomachs and abomasums for position and adhesions
 - Remove forestomachs and abomasums as a unit; separate serosal attachments to stretch the organs out. Open and examine each organ. Remove ingesta and rinse the rumen mucosa with water to examine.

Guidelines for Packing and Shipping of Samples

1. Label all sample containers with the following information, using indelible ink:
 - Client name
 - Animal name
 - Case number (if used)
 - Date of collection
 - Site of collection (e.g. liver, right kidney)
2. Ship in plastic containers whenever possible
3. Be sure that lids are tight on containers that contain liquid. The ratio of formalin to soft tissue should be 10:1. If the sample is bone, the ratio should be 20:1.
4. Containers with liquid should be placed into zippered plastic bags separate from submission forms to prevent forms from becoming damaged and unreadable in case of leaks
5. Complete submission forms, including all requested information
6. Include address, phone number and FAX number for your clinic to facilitate return of results
7. Pack container and submission form in box for mailing, allowing room for packing materials such as foam peanuts, bubble plastic, or newspaper as appropriate
8. Check with lab to determine if sample must be received at room temperature, cool, or frozen and include coolant source if needed

CLINICAL MENTORSHIP TASKS

INTRODUCTION TO ESSENTIAL TASKS AND CRITERIA

Before starting each task:

1. Read the Goal, Description, Criteria, and Materials to be submitted for Evaluation and Verification. Understand what is expected of you for each task.
2. Make sure you have whatever equipment you need to document the task. Pay particularly close attention to the details of what needs to be documented.
3. Make sure you obtain appropriate permissions where necessary. Please keep the facility's owner/manager aware of your activities. A continued good relationship with the veterinarian in charge is key to having a positive Clinical Mentorship experience.
4. Label documentation so it is easy to find all components of the materials you submit for evaluation and validation at Purdue.
5. Once everything is completed, package all the paper, video, photographic or other required documentation and send it to the VT-DLP at the address provided in the *Completion of Clinical Mentorship* section above.

Assist in Prosection on Non-Preserved Animal

Goal: To assist a DVM in performing basic prosection techniques on a non-preserved animal for purposes of necropsy and sample collection.

Description: The student will assist a DVM in performing basic prosection techniques on a non-preserved animal, identifying internal structures as they are exposed.

Criteria: The student checked and recorded signalment for the patient.

The student reviewed the patient history and clinical diagnosis.

The student reviewed laboratory data submitted with the animal.

The student examined and recorded the patient's external appearance, including

- body condition
- mucous membranes
- body orifices
- general conformation
- superficial lesions (tumors, dermatitis, etc.)
- hair coat
- parasites
- lips, gums, cheeks and teeth

*Note: External examination should be done with the DVM present, or the DVM should examine the animal before proceeding.

The student placed the animal in left lateral recumbency.

The student correctly identified the following structures during the prosection:

- exposed superficial lymph nodes
- jugular veins
- mammary glands or testes
- prepuce and penis (male animal)
- diaphragm
- larynx
- tonsils
- esophagus
- trachea

- lungs
- heart
- liver
- gall bladder
- stomach and pyloric region
- mesentery
- kidneys
- spleen
- small intestine
- large intestine
- urinary bladder

The student collected initial samples for microbiology as directed by the DVM.

The student accurately documented all observations made by the DVM.

Number of Times Task Needs to be Successfully Performed: 1

Materials Submitted for Evaluation and Verification:

1. Summary evaluation form for the Assisting in Prosection on a Non-Preserved Animal task, signed by the clinical mentorship supervisor.
2. One videotape showing the student assisting with a prosection procedure. The tape will clearly show all criteria for the task, and the student will narrate the procedure, including identification of required structures.
3. Copy of written documentation of findings by student and DVM during the prosection.

Summary Evaluation Form for Assisting in Prosection on Non-Preserved Animal

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
 DVM, VMD

Date completed _____

Patient _____

	Unsatisfactory	Satisfactory	Excellent
The student checked and recorded signalment for the patient	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student reviewed the patient history and clinical diagnosis	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student reviewed laboratory data submitted.	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student examined and recorded the patient's external appearance, including			
• body condition	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• mucous membranes	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• body orifices	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• general conformation	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• superficial lesions (tumors, dermatitis, etc.)	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• hair coat	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• parasites	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• lips, gums, cheeks and teeth	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
 *Note: External examination should be done with the DVM present, or the DVM should examine the animal before proceeding			
The student placed the animal in left lateral recumbency	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Unsatisfactory Satisfactory Excellent

The student correctly identified the following structures during the prosection:

- | | | | |
|-----------------------------------|--------------------------|--------------------------|--------------------------|
| • exposed superficial lymph nodes | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • jugular veins | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • mammary glands or testes | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • prepuce and penis (male animal) | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • diaphragm | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • larynx | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • tonsils | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • esophagus | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • trachea | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • lungs | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • heart | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • liver | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • gall bladder | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • stomach and pyloric region | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • mesentery | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • kidneys | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • spleen | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • small intestine | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • large intestine | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |
| • urinary bladder | <input type="checkbox"/> | <input type="checkbox"/> | <input type="checkbox"/> |

The student collected initial samples for microbiology as directed by the DVM

The student accurately documented all observations made by the DVM

Signature of the Clinical Mentorship Supervisor _____

Collection, Preservation and Shipping of Samples

Goal: To collect, preserve, and ship samples collected during a necropsy procedure.

Description: The student will collect samples of liver, kidney and intestine during a necropsy procedure, and properly preserve and ship the samples for histopathologic evaluation according to laboratory and practice protocols.

Criteria: The student examined the liver grossly and collected samples of lesions and adjacent normal tissue, ½ cm thick and 1cm X 1cm square. If the liver was grossly normal, the student collected a representative sample (½ X 1 X 1cm).

The student examined the kidneys grossly and collected samples of lesions and adjacent normal tissue in longitudinal wedge sections ½ cm thick. If the kidneys were grossly normal, the student collected a representative sample (½ cm longitudinal wedge).

The student examined the intestines grossly and collected samples of lesions and adjacent normal tissue in ½ cm cross-sections. Care was taken not to touch the mucosal surface, and the sample was rinsed gently under water to remove ingesta and feedstuffs. If the intestine was grossly normal, the student collected a representative sample (½ cm cross-section).

The student placed the tissue samples into separate formalin jars, one jar per site / organ. The kidney samples are separated by left or right kidney.

The student labeled each formalin jar with the following information:

- Client name
- Animal name
- Case number (if used)
- Date of collection
- Site of collection (e.g., liver, right kidney)

The student completed submission forms for samples collected.

The student properly packaged the samples for shipping.

Number of Times Task Needs to be Successfully Performed: 1

Materials Submitted for Evaluation and Verification:

1. Summary evaluation form for the Collection, Preservation and Shipping of Samples task, signed by the clinical mentorship supervisor.
2. One videotape showing the student collecting samples and properly preserving and packaging them. The tape will clearly show all criteria for the task, including labels on jars.
3. Copy of submission form completed by the student to be sent with samples to a laboratory (even if samples will not actually be sent to the lab).

Summary Evaluation Form for Collection, Preservation and Shipping of Samples

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
 DVM, VMD

Date completed _____

Patient _____

	Unsatisfactory	Satisfactory	Excellent
The student examined the liver grossly and collected samples of lesions and adjacent normal tissue, ½ cm thick and 1cm X 1cm square. If the liver was grossly normal, the student collected a representative sample (½ X 1 X 1cm).	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student examined the kidneys grossly and collected samples of lesions and adjacent normal tissue in longitudinal wedge sections ½ cm thick. If the kidneys were grossly normal, the student collected a representative sample (½ cm longitudinal wedge).	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student examined the intestines grossly and collected samples of lesions and adjacent normal tissue in ½ cm cross-sections. Care was taken not to touch the mucosal surface, and the sample was rinsed gently under water to remove ingesta and feedstuffs. If the intestine was grossly normal, the student collected a representative sample (½ cm cross-section).	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student placed the tissue samples into separate formalin jars, one jar per site / organ. The kidney samples are separated by left or right kidney.	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student labeled each formalin jar with the following: <ul style="list-style-type: none"> • Client name • Animal name • Case number (if used) • Date of collection • Site of collection (e.g., liver, right kidney) 	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student completed submission forms for samples collected.	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
The student properly packaged the samples for shipping.	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Signature of the Clinical Mentorship Supervisor _____

Safe Handling of Rabies Suspect Bodies and Samples Project

The student will provide a written paper detailing the clinic SOP for handling of rabies suspect bodies. The first section will be for a patient that needs to be tested for rabies only. The second section will be for a patient that needs a necropsy performed but is a rabies suspect. There will also be a section on human exposure to rabies.

Section 1 – Testing Animal for Rabies Only

Address the following:

1. Describe your clinic SOP for a stray dog of unknown history brought to your clinic for euthanasia after biting a human. Include how clinic personnel protect themselves from possible exposure to rabies.
2. Describe how the dog is submitted for rabies testing, including how the brain is collected, preserved and submitted.
3. Evaluate the above protocol and describe changes that you would make in the protocol, and why.

Section 2 – Necropsy on Rabies Suspect

Address the following:

1. Describe your clinic SOP for a dog that was a patient in your clinic, was a rabies suspect but had not bitten anyone, and died in your clinic. The cause of death is unknown and the owner requests a necropsy. Include how clinic personnel protect themselves from possible exposure to rabies.
2. Describe how the dog is submitted for rabies testing, as well as how the necropsy is accomplished safely.
3. Evaluate the above protocol and describe changes that you would make in the protocol, and why.

Section 3 – Human Exposure to Rabies

Discuss what action should be taken if a patient that was treated at your hospital tests positive for rabies. What should a human healthcare provider recommend for the staff who handled the patient? For more information, visit the website for the Center for Disease Control at <http://www.cdc.gov/> and look for rabies under “Health Topics A-Z”.

Storage and Disposal of Dead Animals Project

The student will provide a written paper detailing the clinic SOP for storage and handling of dead animals. The following questions should be addressed:

1. Describe storage of dead animals that are for disposal only, not to be necropsied. Include storage “containers” used and where the storage area is located. Discuss labeling methods.
2. Describe storage of dead animals that require necropsy. Include storage “containers” used and where the storage area is located. Discuss labeling methods.
3. Describe disposition options available to an owner whose pet has died (cremation, burial, return body to owner, etc.). Include storage and labeling methods during the holding period until disposition occurs.
4. Describe the clinic SOP for disposal of dead animals that do not have owners or whose owners do not have a preference for how the body is disposed. Include storage and labeling methods during the holding period until disposition occurs.
5. Evaluate the clinic’s protocol for the handling of dead animals. Describe changes that you would make in the protocol, and why.