

Brief Communication

Potassium channel blocker, 4-aminopyridine-3-methanol, restores axonal conduction in spinal cord of an animal model of multiple sclerosis

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ABSTRACT

Multiple sclerosis (MS) is a severely debilitating neurodegenerative diseases marked by progressive demyelination and axonal degeneration in the CNS. Although inflammation is the major pathology of MS, the mechanism by which it occurs is not completely clear. The primary symptoms of MS are movement difficulties caused by conduction block resulting from the demyelination of axons. The possible mechanism of functional loss is believed to be the exposure of potassium channels and increase of outward current leading to conduction failure. 4-Aminopyridine (4-AP), a well-known potassium channel blocker, has been shown to enhance conduction in injured and demyelinated axons. However, 4-AP has a narrow therapeutic range in clinical application. Recently, we developed a new fast potassium channel blocker, 4-aminopyridine-3-methanol (4-AP-3-MeOH). This novel 4-AP derivative is capable of restoring impulse conduction in *ex vivo* injured spinal cord without compromising the ability of axons to follow multiple stimuli. In the current study, we investigated whether 4-AP-3-MeOH can enhance impulse conduction in an animal model of MS. Our results showed that 4-AP-3-MeOH can significantly increase axonal conduction in *ex vivo* experimental autoimmune encephalomyelitis mouse spinal cord.

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Introduction

It is well documented that myelin damage could expose potassium channels at juxtaparanodal regions, which are covered by myelin in healthy axons (Chiu and Ritchie, 1980; Sherratt et al., 1980; Haghghi et al., 1995; Nashmi et al., 2000; Karimi-Abdolrezaee et al., 2004; Sun et al., 2010). The potassium efflux through fast potassium channels is believed to cause axonal conduction block by preventing sufficient depolarization and initiation of the action potential at the node of Ranvier. Consistent with this notion, 4-AP, a known potassium channel blocker, has been shown to restore action potential conduction in spinal cord injury where demyelination is a major pathological phenomenon (Blight, 1985, 1989; Blight et al., 1991; Waxman, 1992; Hayes et al., 1994; Waxman et al., 1994; Shi and Blight, 1997; Shi et al., 1997; Jensen

and Shi, 2003; Totoiu and Keirstead, 2005). The significance of 4-AP-mediated axonal conduction restoration was further highlighted by the recent approval of this drug by the FDA to improve walking in multiple sclerosis (MS) patients (Ampyra, Acorda Therapeutics, Inc.).

However, 4-AP has a narrow therapeutic range with side effects including dizziness, nausea, and seizures (Stork and Hoffman, 1994; Potter et al., 1998; Pena and Tapia, 1999, 2000; Korenke et al., 2008). In an effort to produce an alternative therapy that targets the fast potassium channels, we have developed a new potassium channel blocker, 4-aminopyridine-3-methanol (4-AP-3-MeOH). This compound has been shown to block fast potassium channels and restore impulse conduction in mechanically injured spinal cord where myelin damage is evident (Sun et al., 2010). Interestingly, the lowest effective concentration of 4-AP-3-MeOH in stretched spinal cords is between 0.01 and 0.1 μM and the lowest effective concentration of 4-AP in compression injury is between 0.1 and 1 μM (Shi et al., 1997; Sun et al., 2010). However, 4-AP-3-MeOH has not been examined in animal models of MS where myelin damage is caused by non traumatic biochemical events (Compston and Coles, 2008). In the current study, we plan to determine whether 4-AP-3-MeOH can enhance axonal conduction in the EAE mouse, an animal model of MS with diffuse myelin damage and associated motor deficits (Gold et al., 2006). Our study demonstrated that 4-AP-3-MeOH can significantly enhance axonal conduction in *ex vivo* EAE mouse spinal cord.

Abbreviations: MS, multiple sclerosis; 4-AP, 4-aminopyridine; 4-AP-3-MeOH, 4-aminopyridine-3-methanol; EAE, experimental autoimmune encephalomyelitis; CAP, compound action potential.

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Experimental procedures

EAE mice were established using an MOG₃₅₋₅₅/CFA emulsion kit followed by pertussis toxin injections (EK-0115, Hooke Laboratories) (Kalyvas and David, 2004). The emulsion was injected subcutaneously in the back of C57BL/6 mice that were 9–10 weeks old. An intraperitoneal injection of pertussis toxin was administered after the emulsion and a second dose of pertussis toxin followed 22–24 h later. The behavior of the mice was assessed daily for 30 days and the symptoms of EAE began to emerge approximately 12–15 days after the emulsion injection. The average score of EAE mice used in this study was 2.2 ± 0.5 ($n = 12$), based on a well-established 5-point behavioral scoring system (Kalyvas and David, 2004). The 5-point scale begins at 0, which represents a healthy animal without any deficit. The symptoms will progress from a limp tail to weakness in the hind limbs and then finally hind limb paralysis. These experiments

were approved by the Purdue Animal Care and Use Committee, West Lafayette, Indiana.

After confirmation of the behavioral deficits, the mice were anesthetized with a combination of Ketamine (90 mg/kg) and Xylazine (10 mg/kg) and then perfused with a cold, oxygenated Krebs' solution. The spinal column was quickly removed from the body and the spinal cord then isolated as previously described (Shi and Blight, 1996; Sun et al., 2010). The samples were incubated in continuously oxygenated Krebs' solution (Shi and Blight, 1996; Sun et al., 2010) before recording. Whole mouse spinal cord (around 3.5 cm long) was placed into the double sucrose-gap recording chamber, allowing the white matter tracts to extend across the apparatus.

The amplitude of the compound action potential (CAP) was recorded from isolated EAE mouse spinal cord using a double sucrose-gap recording system, which is similar to that described previously (Shi and Blight, 1996; Jensen and Shi, 2003; Shi and Whitebone,

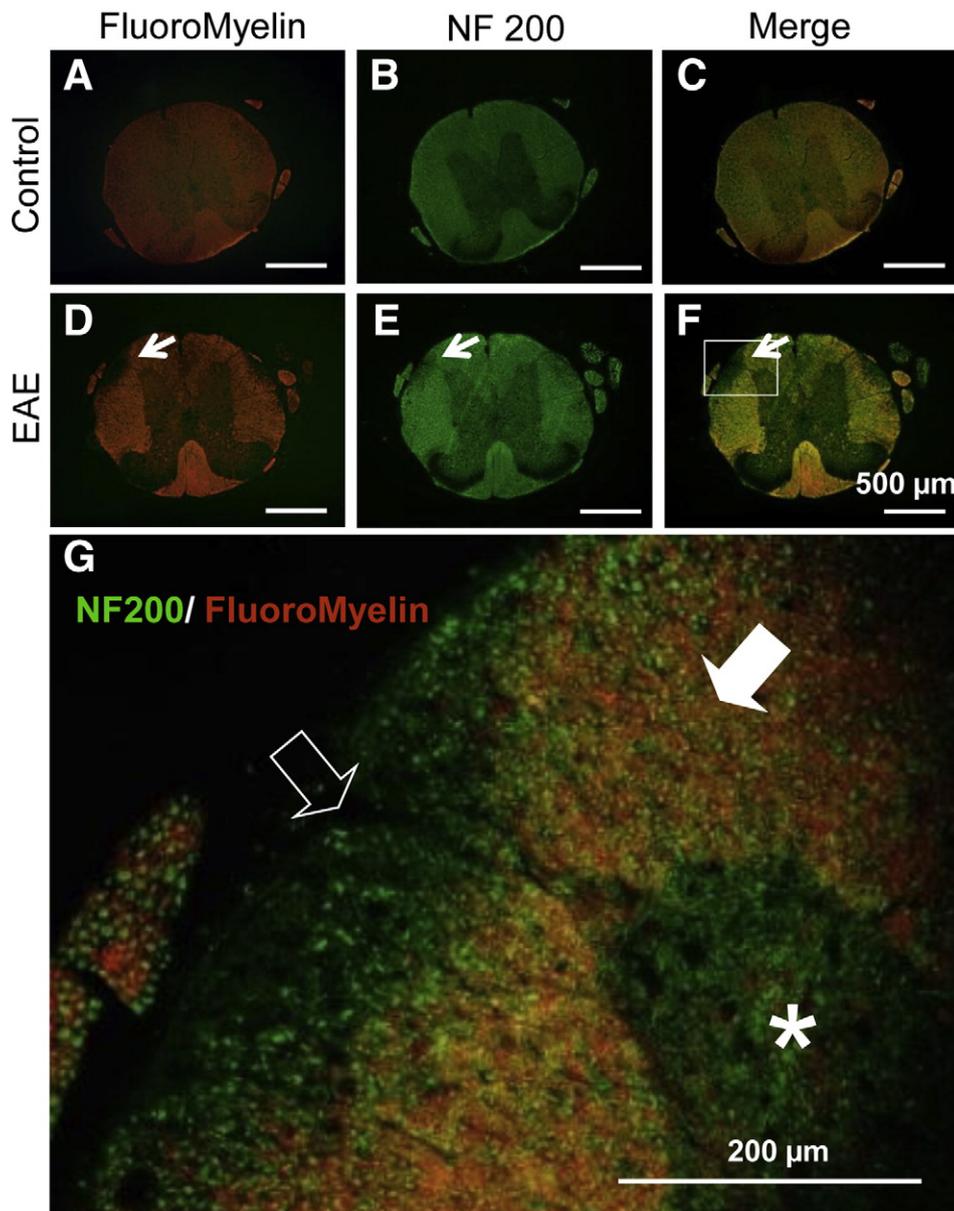


Fig. 1. Images of thoracic cross-sections of spinal cords from control (A–C) and EAE (D–F) mice labeled with fluoromyelin (red) and anti-NF200 (green). No noticeable demyelinated areas were found in the white matter of control mice spinal cord (A–C). In EAE mouse spinal cord sections, a typical demyelinated region in the white matter is shown (arrow) (D–F). (G) A photomicrograph displays the higher magnification of the region circumscribed in F. Note that many axons are labeled without corresponding myelin staining. The open arrow indicates area where axons lack myelin, and filled arrow denotes area of axons with normal myelin appearance. Asterisk indicates gray matter of spinal cord. Scale bars = 500 μm (A–F); 200 μm (G).

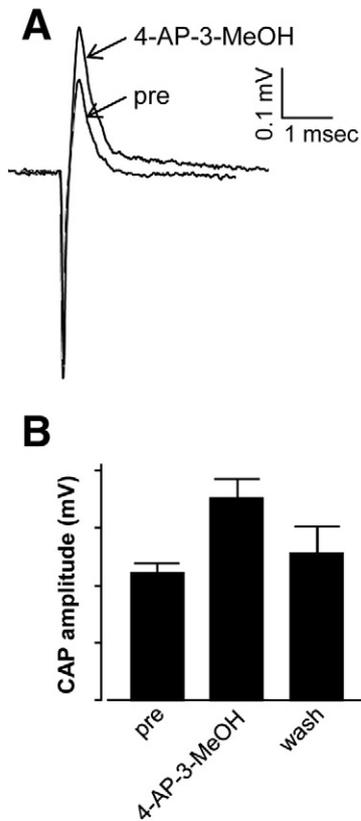


Fig. 2. The electrophysiological response of *ex vivo* EAE mouse spinal cord to the application of 100 μ M 4-AP-3-MeOH showed an increased in CAP amplitude. Representative CAPs of pre and post 4-AP-3-MeOH treatment were superimposed and shown in A. Graph (B) demonstrates that 4-AP-3-MeOH treatment significantly enhanced CAP amplitude from 0.11 ± 0.01 mV to 0.17 ± 0.02 mV ($n = 5$, $P < 0.05$). The CAP amplitude returned to 0.13 ± 0.02 mV following wash for 45–60 min. Error bars represent SEM.

2006). Baseline CAP measurements of EAE spinal cord were recorded before applying 4-AP-3-MeOH. 100 μ M of fresh 4-AP-3-MeOH was then applied through Krebs' circulation in the central well to the spinal cord. Lastly, the spinal cord was then washed with Krebs' solution. The temperature was maintained at 37 $^{\circ}$ C.

For immunofluorescence staining, mice were perfusion fixed and then the whole spinal cord was extracted as previously mentioned

(Shi and Blight, 1996). Three EAE mice were used as well as three control mice. Subsequently, 15 μ m spinal cord cross-sections were cut and labeled with Fluoromyelin (Invitrogen, CA) and Anti-Neurofilament 200 antibody produced in rabbit (dilution ratio 1:150, Sigma-Aldrich). Alexa Fluor[®] 488 goat anti-rabbit IgG (H + L) was used as a secondary antibody (dilution ratio 1:100, Invitrogen, CA).

Results

Fluoromyelin (myelin sheath) and anti-NF200 (axons) were used to define areas of demyelination in spinal cord cross-sections at the thoracic level. Areas with low or no Fluoromyelin were noted in EAE mice, indicating the region of demyelination, typical of multiple sclerosis (Gold et al., 2006). No noticeable demyelinated areas were found throughout the thoracic region in control cords (Fig. 1).

The effect of 4-AP-3-MeOH on axonal conduction in EAE mice was examined using extracted spinal cord placed in a double sucrose-gap recording chamber. The CAP amplitude before drug treatment was 0.11 ± 0.01 mV ($n = 5$). 4-AP-3-MeOH at 100 μ M significantly improved CAP to 0.17 ± 0.02 mV following a 45-min application, representing a 55% increase in amplitude ($P < 0.05$, $n = 5$). The CAP amplitude returned to 0.13 ± 0.02 mV following wash for 45–60 min (Fig. 2B). In contrast, 4-AP-3-MeOH did not cause significant changes in CAP amplitude in control healthy mice. Specifically, CAP amplitudes for pre-drug, drug, and wash are 0.285 ± 0.04 , 0.298 ± 0.06 , and 0.285 ± 0.06 mV, respectively ($n = 3$ for all groups, $P > 0.5$).

When a series of stimulus voltages with intensities ranging from 1.85 to 6.5 V were applied before and during the presence of 4-AP-3-MeOH in EAE mice, it is clear that the increase of CAP amplitude as a result of 4-AP-3-MeOH was evident in a wide range of stimulus intensities (Fig. 3A). In addition, a comparison of the response amplitude, with and without 4-AP-3-MeOH, showed no significant differences in the relative increase of amplitude (Fig. 3A). This relationship is further demonstrated in absolute terms in Fig. 3B. The near unity of the slope indicates that 4-AP-3-MeOH-mediated CAP conduction was not biased based on axonal caliber and threshold (Fig. 3B).

Discussion

Similar to our prior study in mechanically injured spinal cord, we have shown in this study that 4-AP-3-MeOH can also significantly enhance axonal conduction in the spinal cord of EAE mice (Fig. 2B). Since 4-AP-3-MeOH has been shown to block fast potassium channels

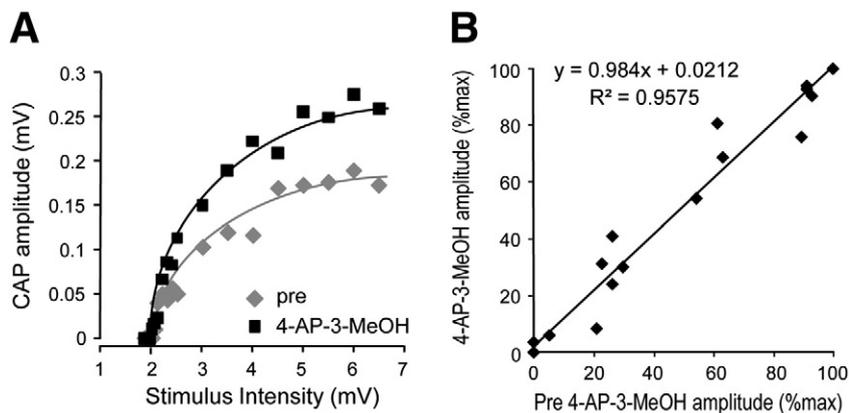


Fig. 3. Assessment of relation between stimulus intensity and response amplitude before and after treatment with 4-AP-3-MeOH. Stimulus intensities ranging from 1.85 to 6.5 V were applied to EAE spinal cord prior to drug exposure and at 100 μ M 4-AP-3-MeOH-treated conditions. (A) Comparison of these two conditions is presented as a graphic plot, with trend lines added for the two conditions. Clearly the response amplitude increased with 4-AP-3-MeOH at nearly all the stimulus intensities. (B) Normalized CAP responses (percentage of maximum CAP in each condition) were plotted as pre-drug against 4-AP-3-MeOH-treated (100 μ M). Original data in B are the same as in A. Overall trends demonstrated a linear correlation between pre-drug and the 4-AP-3-MeOH-treated group, indicating that there is little difference in the sensitivity of axons with different stimulus thresholds to 4-AP-3-MeOH-mediated amplitude enhancement.

(Sun et al., 2010), it is likely that 4-AP-3-MeOH enhances CAP by blocking this type of channel unmasked due to myelin damage. To our knowledge, this is the first time that a potassium channel blocker other than 4-AP has been shown to enhance axonal conduction in an animal model of MS.

Based on available evidence, there are two major differences between 4-AP-3-MeOH and 4-AP. First of all, the lowest concentration of 4-AP-3-MeOH that can produce significant axonal conduction restoration is between 0.01 to 0.1 μM while 4-AP is between 0.1 and 1 μM (Shi and Blight, 1997; Shi et al., 1997; Sun et al., 2010). Second, unlike axons affected by 4-AP, axons affected by 4-AP-3-MeOH conduct action potentials in a manner similar to normal axons based on *ex vivo* examinations (Jensen and Shi, 2003; Sun et al., 2010). Specifically, while producing a significant conduction recovery, 4-AP (100 μM) induced an increase of refractoriness. On the other hand, 4-AP-3-MeOH at the same concentration did not affect the axonal refractoriness while inducing conduction recovery (Jensen and Shi, 2003; Sun et al., 2010). However, caution should be taken when extrapolating these data to *in vivo* situations where the safe effective concentration could be hundreds of times lower than used *ex vivo* (Shi and Blight, 1997; Jensen and Shi, 2003; Goodman et al., 2009; Sun et al., 2010). Similar to that in 4-AP, our data indicate that axons with various axonal diameters benefit equally from 4-AP-3-MeOH-induced axonal conduction restoration.

Based on these findings using *ex vivo* testing, it appears that 4-AP-3-MeOH could potentially be an alternative therapeutic option in addition to 4-AP in enhancing axonal conduction in demyelinated axons. In the current study, we chose the most beneficial concentration determined in *ex vivo* traumatic spinal cord injury study to determine its effectiveness in EAE mouse spinal cord. Further experiments testing a range of concentrations are necessary to establish a dose response for 4-AP-3-MeOH-mediated axonal conduction restoration in EAE mice.

Since these are the initial findings of 4-AP-3-MeOH on isolated spinal cord extracted from EAE mice, there are still more possible differences compared to 4-AP when applied *in vivo*. Specifically, future studies *in vivo* will need to assess various factors that could affect the concentration achieved in CNS, which include metabolism, absorption, and excretion of 4-AP-3-MeOH. These factors are crucial to determine the eventual efficacy, potency, and side effects and therefore clinical value of 4-AP-3-MeOH in the treatment of MS patients.

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