

# SAMPLE SUBMISSION GUIDELINES FOR THE CLINICAL PATHOLOGY LABORATORY

The following guidelines should be used when collecting samples for submission the next day.

Sample should be shipped via overnight/next day delivery on an ice pack with completed submission form.

Please do not ship samples on Fridays as there is no mail service at the University on Saturdays.

### **HEMATOLOGY**

# Sample Type:

- One EDTA (lavender top) tube at least ½ full. Mix sample well immediately after collection.
   Sample must be free of clots.
- 2. Two freshly made unstained, unfixed blood smears.

#### **Additional Comments:**

- Smears should be made from a well-mixed EDTA sample immediately after collection. Smears
  made in the laboratory from courier samples may not be adequate if sample is over 8 hours old.
- 2. Smear Preparation technique aids:
  - a. Use clean glass slides. \*\*No re-used slides\*\*
  - b. Place a drop of blood ~3mm in diameter at one end of the slide. Note a drop of blood too small or large will create a long or short smear.

c. Hold pusher slide at a 30° - 45° angle.

# **CHEMISTRY (Routine)**

#### Sample Type:

1. 1 - 2 mL of serum from a red top tube

#### **Additional Comments:**

- 1. Allow the blood to clot for 30 minutes and then centrifuge at a high speed for 10 minutes.
- 2. Remove the serum from the clot tube and transfer it into a no-additive tube (another red top tube is acceptable. Clearly mark tube with patient ID and SERUM.
- The most frequent causes of inaccurate biochemical tests are aged sample, hemolysis, icterus, lipemia, and insufficient sample quantity.
- 4. Serum should be stored in the refrigerator prior to shipping, with the exception of Fructosamine.
- 5. Store and ship serum for Fructosamine frozen, on dry ice, for next day delivery. (\*Hemolysis is not recommended for fructosamine testing. Submit only non-hemolyzed serum for fructosamine testing.)



# **COAGULATION**

Call our lab prior to collecting sample to ensure we can assist you with this test. Sample stability is very limited.

#### **URINALYSIS**

#### Sample Type:

10 mL of urine in a sterile leak-proof container

#### **Additional Comments:**

- 1. Urine testing should not be delayed.
- Urine should be refrigerated immediately after collection to delay any chemical and cytological changes of the urine.

# **MISCELLANEOUS TEST**

Instructions: Please contact the laboratory for sample collection and shipping instructions.

- 1. Endogenous ACTH
- 2. Ammonia
- 3. Coombs
- 4. Avian samples
- 5. Coagulation Tests



#### **CYTOLOGY**

- ALL slides/smears should be labeled with patient name, lesion location and date.
- Place smears in a slide box prior to shipping. No ice pack required when shipping slides only.
- Ship samples/smears on an ice pack overnight (avoid direct contact between the sample and ice pack).
- A. Fluid (Synovial, Tracheal, etc.)

Submit:

- 1. Primary tube for Sample is an EDTA (lavender top tube).
- 2. Two air-dried (unfixed, unstained) smears of the sample.
- B. CSF Fluid

Submit:

- 1. Primary tube for sample is RED top tube. This will allow us to run a micro-protein on the sample if sample quantity allows. Secondary tube for sample is EDTA (lavender top tube).
- 2. Two air-dry (unfixed, unstained) smears of the sample
- C. Aspirate and Impression smears

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#### Submit:

1. Air-dried (unfixed, unstained) smears.

# **SAMPLE SUBMISSION REMINDERS:**

- 1. Always label all your samples with the patient's name and date.
- 2. Smears should be made on a clean frosted slide.
- 3. Always make sure sample(s) are in spill proof containers.
- 4. EDTA whole blood, serum, urine, and cytology samples should be shipped on ice packs. Avoid direct contact between the sample(s) and ice pack.
- 5. Always make sure smears are placed in a slide box to avoid breakage.
- 6. If you read your own slides, please include at least 1 unstained, unfixed slide before sending.
- 7. Always ship samples overnight and do not ship on Fridays. We DO NOT receive samples on weekends.

# **Ship Samples To:**

Clin Path Lab 625 Harrison St Lynn Hall G351

West Lafayette, IN 47907



If you have any questions, please feel free to contact the lab.

765-494-7563 or 765-494-3972

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